


 Cite this: *RSC Adv.*, 2020, **10**, 3212

# Confirmatory non-invasive and non-destructive differentiation between hemp and cannabis using a hand-held Raman spectrometer†

 Lee Sanchez,<sup>a</sup> Conor Filter,<sup>b</sup> David Baltensperger<sup>c</sup> and Dmitry Kurouski<sup>id</sup>\*<sup>ad</sup>

Cannabis is a generic term that is used to denote hemp plants (*Cannabis sativa*) that produce delta-9-tetrahydrocannabinolic acid (THCA) in amounts higher than industrial hemp. While THCA itself is not considered psychoactive, it is the source of the psychoactive delta-9 tetrahydrocannabinol (THC) that forms from its oxidation. About 147 million people, which is around 2.5% of the world population, consume cannabis. This makes cannabis by far the most widely cultivated and trafficked illicit drug in the world. Such enormous popularity of cannabis requires substantial effort by border control and law enforcement agencies to control illegal trafficking and distribution. Confirmatory diagnostics of cannabis is currently done by high pressure liquid chromatography (HPLC), which requires sample transportation to a certified laboratory, making THC diagnostics extremely time and labor consuming. This catalyzed a push towards development of a portable, confirmatory, non-invasive and non-destructive approach for cannabis diagnostics that could be performed by a police officer directly in the field to verify illicit drug possession or transport. Raman spectroscopy (RS) is a modern analytical technique that meets all these strict expectations. In this manuscript, we show that RS can be used to determine whether plant material is hemp or cannabis with 100% accuracy. We also demonstrate that RS can be used to probe the content of THCA in the analyzed samples. These findings suggest that a hand-held Raman spectrometer can be an ideal tool for police officers and hemp breeders to enable highly accurate diagnostics of THCA content in plants.

 Received 9th October 2019  
 Accepted 27th December 2019

DOI: 10.1039/c9ra08225e

[rsc.li/rsc-advances](http://rsc.li/rsc-advances)

## Introduction

Drug trafficking involves cultivation, manufacture, distribution and sale of substances that are subject to drug prohibition laws.<sup>1</sup> On a federal and state-specific level, drug trafficking is considered a felony offense that carries serious consequences such as prison time and significant monetary fines. Worldwide, sentences for drug distribution and trafficking generally range from 3–5 years to life in prison or even the death penalty.<sup>1</sup>

Cannabis is by far the most widely cultivated and trafficked illicit drug in the world.<sup>1</sup> The problem of cannabis trafficking is especially challenging in the United States, as 11 states and Washington DC have legalized it for recreational use, 33 states have legalized cannabis for medicinal use and 15 states have

decriminalized cannabis.<sup>2</sup> U.S. Customs and Border Protection, the Drug Enforcement Administration, and state and local police departments across the country spend enormous financial resources to cease the traffic of illicit drugs. For instance, in 2015 alone, the federal government spent an estimated \$9.2 million every day towards the incarceration of people charged with drug-related offenses; and since 1971, the war on drugs has cost the United States an estimated \$1 trillion.<sup>3</sup>

A substantial portion of these expenses is used for forensic analyses of potential drug substances, which are primarily done by HPLC and mass spectrometry.<sup>4–7</sup> These sophisticated tests are destructive, time consuming and can only be performed in certified laboratories. This drastically delays the times of analysis for potential drug substances.<sup>7</sup> This problem has catalyzed a push towards the development of portable tests that can be performed directly in the field and can confirm the presence of an illicit drug. Several companies have come up with tests that were based on a color change upon interaction of a specific reagent with the drug of interest. However, such tests did not find broad applications in forensic practice due to their high cost, destructive nature and a lack of a quantitative response.<sup>8</sup> Their application is far more challenging for hemp vs. cannabis diagnostics because hemp may legally contain up to 0.3% THC, the major psychoactive component of cannabis. However, this

<sup>a</sup>Department of Biochemistry and Biophysics, Texas A&M University, College Station, Texas 77843, USA. E-mail: dkurouski@tamu.edu

<sup>b</sup>Texas Farm & Process, LLC, Fort Worth, Texas 76102, USA

<sup>c</sup>Department of Soil and Crop Sciences, Texas A&M University, College Station, Texas 77843, USA

<sup>d</sup>The Institute for Quantum Science and Engineering, Texas A&M University, College Station, Texas, 77843, USA

† Electronic supplementary information (ESI) available: HPLC results of cannabis analyses and Fig. S1 and S2. See DOI: 10.1039/c9ra08225e



legal amount of THC can give positive color change on those tests.

We hypothesized that hemp *vs.* cannabis diagnostics can be done using Raman spectroscopy. This analytic technique is based on inelastic light scattering of photons, which excite molecules in the sample to higher vibrational or rotational states.<sup>9</sup> After these inelastically scattered photons are collected by a spectrometer, the change in the photon energy is determined. Since the change in the photon energy will directly depend on the vibrational properties of the sample, RS can be used to probe structure and composition of analyzed specimen. Our group previously demonstrated that RS can be used to detect and identify presence of urine on police uniform<sup>10</sup> as well as diagnose biotic and abiotic stresses on plants.<sup>11,12</sup>

In the current study, we show that a hand-held Raman spectrometer can be used to determine whether the sample of interest is hemp or cannabis with 100% accuracy. Moreover, we show that RS can be used to probe the content of THCA in cannabis. This makes this approach highly suitable for police as such analysis is non-invasive and non-destructive and can be performed directly in the field.

## Experimental

### Plants

Hemp and cannabis plants were grown at Evergreen Enterprises LLC located in Denver, CO. Buds of 5–10 fresh hemp plants and 10–15 different fresh-frozen cannabis plants of three varieties known as “triple chocolate chip (TCC)”, “gelato cake (GC)”, and “twisted sherbert (TS)” were used. Fresh-freezing of plants was performed by placement of plant buds into freezer at  $-10$  to  $-15$  °C. Fresh-freezing is a standard procedure in cannabis farming that is used to preserve cannabinol content of plants during their post-harvest processing. Based on visual examination, fresh-freezing does not result in any noticeable changes in plant appearance or texture. Cannabinol content of hemp (0.07% THCA) and cannabis (TCC: 10.31% THCA, GC: 6.12% THCA, and TS: 4.05% THCA) strains was determined by Agricorn Laboratories. Certificates of analyses are provided in the ESI.†

### Raman spectroscopy

Raman spectra were taken with a hand-held Resolve Agilent spectrometer equipped with 831 nm laser source (beam diameter  $\sim 2$  mm). Resolve spectral resolution was  $15\text{ cm}^{-1}$ . Samples were brought in a direct contact with the spectrometer for spectral acquisition. The following experimental parameters were used for all collected spectra: 10 s acquisition time, 495 mW power. The spectra were automatically baselined by the instrument software. In total, 20–23 spectra were collected from each sample type (hemp or individual cannabis variety). Spectra shown in the manuscript are raw baseline corrected, without smoothing.

### Multivariate data analysis

SIMCA 14 (Umetrics, Umeå, Sweden) was used for statistical analysis of the collected Raman spectra. All imported spectra

were scaled to unit variance to give all spectral regions equal importance. Orthogonal partial least squares discriminant analysis (OPLS-DA) was performed in order to determine the number of significant components and identify spectral regions that best explain the separation between the classes. Standard normal variate (SNV) correction followed by the first derivative was applied to the spectra normalized by the total area. Next, using OPLS-DA, we determined the number of predicting and orthogonal significant components and identified spectral regions that best explain the separation between the classes. In order to give each of the spectral regions equal importance, all spectra were scaled to unit variance. Raw spectra, containing wavenumbers  $701\text{--}1700\text{ cm}^{-1}$ , were retained in the model that resulted from this iteration of OPLS-DA.

## Results and discussion

We found that the Raman spectrum of hemp is dominated by vibrational bands that could be assigned to cellulose, carotenoids and lignin, Fig. 1, Table 1.<sup>13</sup> At the same time, spectra collected from GC, TCC and TS clearly demonstrate presence of THCA. Specifically, we observed vibrational bands at  $780$ ,  $1295$ ,  $1623$ , and  $1666\text{ cm}^{-1}$  in the spectra of GC, TCC and TS that can be assigned to THCA. We also observed a change in the intensities of bands at  $916$ ,  $993$ ,  $1000$ ,  $1084$ ,  $1155$ ,  $1185$ ,  $1267$ ,  $1321$ , and  $1525\text{ cm}^{-1}$  in the spectra collected from GC, TCC and TS relative to the intensities of these bands in the Raman spectrum of hemp. However, the change in intensities of these vibrational bands cannot be used for confirmatory diagnostics of THCA presence because these bands have also been observed in the spectrum of hemp and can be assigned to carbohydrates and cellulose. Nevertheless, the change in intensities of these vibrational bands suggests about structural differences between hemp and cannabis plants. For instance, we found that

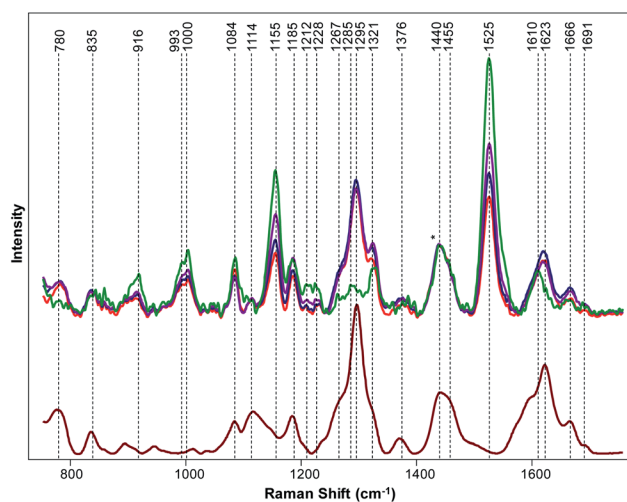


Fig. 1 Top: Raman spectra collected from hemp (green), GC (purple), TCC (blue) and TS (red). Bottom: Raman spectrum of THCA extract (maroon). Spectra normalized on  $\text{CH}_2$  vibrations ( $1440$  and  $1455\text{ cm}^{-1}$ ) that are present in nearly all classes in biological molecules (marked by asterisks (\*)).



Table 1 Vibrational bands and their assignments for hemp, cannabis species and THCA

Band	Vibrational mode	Assignment
780	TBA	THC/THCA
835	TBA	THC/THCA
916	$\nu(\text{C-O-C})$ in plane, symmetric	Cellulose, lignin <sup>14</sup>
993–1000	$\nu_3(\text{C-CH}_3)$ stretching) and phenylalanine	Carotenoids, protein <sup>8,15</sup>
1084	$\nu(\text{C-O}) + \nu(\text{C-C}) + \delta(\text{C-O-H})$	Carbohydrates <sup>16</sup>
1114	$\nu_{\text{sym}}(\text{C-O-C})$ , C-OH bending	Cellulose <sup>17,18</sup>
1155	$\nu_{\text{asym}}(\text{C-C})$ ring breathing	Carbohydrates, cellulose <sup>14</sup>
1185	$\nu(\text{C-O-H})$ next to aromatic ring + $\sigma(\text{CH})$	Xylan <sup>19,20</sup>
1212–1228	$\delta(\text{C-C-H})$	Aliphatic, <sup>21</sup> xylan <sup>19</sup>
1267	C-O stretching (aromatic)	Lignin <sup>22</sup>
1285	$\delta(\text{C-C-H})$	Aliphatic <sup>21</sup>
1295	TBA	THC/THCA
1321	$\delta\text{CH}_2$ bending vibration	Cellulose, lignin <sup>14</sup>
1376	$\delta\text{CH}_2$ bending vibration	Aliphatic <sup>21</sup>
1440	$\delta(\text{CH}_2) + \delta(\text{CH}_3)$	Aliphatic <sup>21</sup>
1455	$\delta\text{CH}_2$ bending vibration	Aliphatic <sup>21</sup>
1527–1551	$-\text{C}=\text{C}-$ (in plane)	Carotenoids <sup>23,24</sup>
1610	$\nu(\text{C-C})$ aromatic ring + $\sigma(\text{CH})$	Lignin <sup>25,26</sup>
1623–	Aromatic	THC/THCA <sup>27</sup>
1691	$\nu(\text{C}=\text{O})$	Carboxyl groups <sup>28</sup>

vibrational bands at 993, 1000 and 1525  $\text{cm}^{-1}$ , which can be assigned to carotenoids, are more intense in the spectrum collected from hemp relative to the intensity of this band in the spectra collected from cannabis species. This result indicate that hemp have higher carotenoid content comparing to cannabis plants. Similarly, we found that intensities of 946, 1084 and 1155  $\text{cm}^{-1}$  bands are stronger in the spectrum of hemp comparing to cannabis plants. This finding suggests that amount of cellulose is higher in hemp comparing to cannabis species.<sup>13</sup>

To further prove our expectation that RS can be used for highly accurate differentiation between hemp and cannabis, we used OPLS-DA analysis. The final model, containing one predictive component, 2 orthogonal components and 1001 (701–1700  $\text{cm}^{-1}$ ) out of 1651 original wavenumbers, was used to generate the misclassification table (Table 2) and the loadings plot (Fig. 2).

The first predictive component (PC) (Fig. 2) explain 94% of the variation between classes. Absolute intensities in the loading spectrum are proportional to the percentage of the total variation between classes explained by each wavenumber. The model identified the peak at 781  $\text{cm}^{-1}$ , which could be assigned to THCA (Table 1), cellulose and lignin peaks at  $\sim 925 \text{ cm}^{-1}$  and the bands at 1260–1320  $\text{cm}^{-1}$ , which correspond to both THCA

and cellulose. Also, the model identified peaks at 1440  $\text{cm}^{-1}$ , which could be assigned to aliphatic vibrations and bands at 1623–1660  $\text{cm}^{-1}$ , which originate from THCA, to be the strongest spectral markers of cannabis, which supports the conclusions of our qualitative spectral analysis above. The model explained 48% of the variation (R2X) in the spectra and correctly assigned all 86 spectra to their classes (Table 2, Fig. S1†). This indicates that coupling of OPLS-DA with RS allows for a 100% accurate differentiation between cannabis and hemp.

Next, we asked a question whether RS can be used for quantitative prediction of the THCA content in cannabis, as well as identification of cannabis variety. HPLC analyses of TCC, GC and TS samples allowed to determine the THCA content which was found to be 10.31%, 6.12% and 4.05%, respectively. Our results demonstrate that intensity of the 1623  $\text{cm}^{-1}$  band directly correlates with the amount of the THCA content in the cannabis, Fig. 1. We used analysis of variance (ANOVA) to visualize the correlation of intensity of 1623  $\text{cm}^{-1}$  band with the content of THCA in the analyzed plant material, Fig. 3. These results suggest that RS can be used for quantitative prediction of the THCA content in intact plant materials. However, more experimental work is needed at this point to determine the

Table 2 Accuracy of classification by OPLS-DA for cannabis (GC, GCC and TS) vs. hemp

	Members	Correct	Cannabis	Hemp
Cannabis	64	100%	64	0
Hemp	22	100%	0	22
Total	86	100%	64	22
Fisher's prob.	$5.9 \times 10^{-21}$			

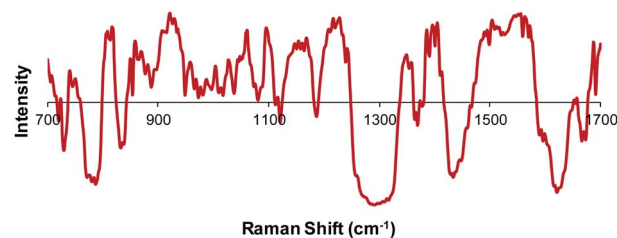


Fig. 2 Loading plot of the predictive component in the Raman spectra of cannabis and hemp.



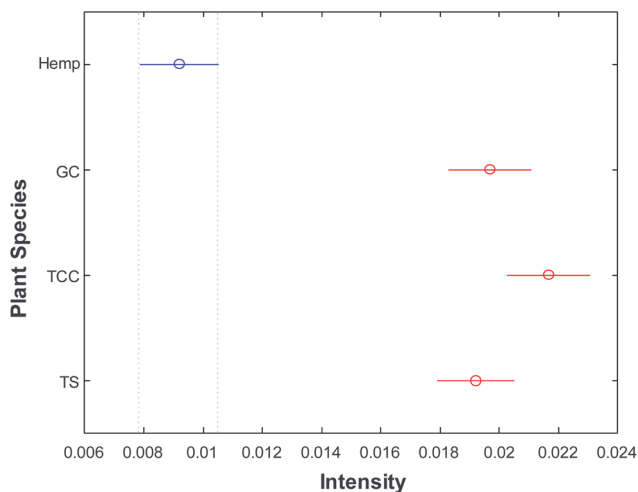


Fig. 3 Means (circles) and confidence intervals for the intensities of 1623 band of THCA for hemp, GC, TCC and TS, normalized to 1440  $\text{cm}^{-1}$ .

accuracy and the range of THCA prediction. This work is currently in progress in our laboratory.

Utilization of OPLS-DA allowed for quantitative differentiation between three different classes on cannabis, Table 3. The final model, containing 2 predictive components, 2 orthogonal components and 1001 (701–1700  $\text{cm}^{-1}$ ) out of 1651 original wavenumbers, was used to generate the misclassification table (Table 3) and the loadings plot (Fig. S2†). The first two predictive components (PC) explain 42% and 36% of the variation between classes respectively, which collectively accounts for 78% of the total class-to-class variation. Absolute intensities in the loadings spectra are proportional to the percentage of the total variation between classes explained by each wave-number within each component. The model identified the peak at 781  $\text{cm}^{-1}$  (PC1), cellulose and lignin peaks at 925  $\text{cm}^{-1}$  (PC1), the bands at 1250–1340  $\text{cm}^{-1}$  (PC1), 1440  $\text{cm}^{-1}$  (PC1) and the region of 1580–1670  $\text{cm}^{-1}$  (PC1) to be the strongest spectral markers representing the three cannabis species, which supports the conclusions of our qualitative spectral analysis above. The model explained 48% of the variation (R2X) in the spectra and 78% (R2Y) of the variation between the classes. Furthermore, the model correctly assigned 84 out of 86 spectra to their classes (Table 3). Our results demonstrate that coupling of OPLS-DA with RS allows for a ~97% accurate identification of the cannabis variety.

Table 3 Accuracy of classification by OPLS-DA for cannabis (GC, GCC and TS) vs. hemp

	Members	Correct	GC	TCC	TS
GC	20	95%	19	1	0
TCC	21	100%	0	21	0
TS	23	96%	1	0	22
Total	64	97%	20	22	22
Fisher's prob.	$1.9 \times 10^{-23}$				

It should be noted that in plants THC is present in carboxylated form of THC, known as THCA. Upon decarboxylation, which can be induced by thermal heating, THCA is converted to THC. Spectroscopically, the carboxyl group of THCA is evident from the vibration at 1691  $\text{cm}^{-1}$ , which was observed in our spectra. One could expect that other vibrational bands that were assigned to THCA would originate from THC since decarboxylation is the only structural transformation in the molecule upon THCA to THC conversion. Specifically, our spectroscopic analysis was based on 1623  $\text{cm}^{-1}$  band, which originate from aromatic moiety present in both THCA and THC. Therefore, we can speculate that RS allows to predict the amount of THC in the analyzed sample without necessary oxidation of THCA to THC.

## Conclusions

Our results clearly demonstrate that RS can be used for confirmatory, non-invasive and non-destructive detection and identification of cannabis. We showed that RS allowed for highly accurate differentiation between cannabis and hemp and prediction of cannabis variety. Because of the portable nature of our analysis, this spectroscopic approach will be highly advantageous for police and border control officers. Also, these results demonstrate that RS can be a great tool for hemp cultivation and breeding allowing for accurate detection of THCA levels in intact growing plants.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

This study was supported by funds from Texas A&M AgriLife Research, Texas A&M University Governor's University Research Initiative (GURI) grant program of (12-2016/M1700437). The authors are also grateful to Charles Farber for the help with ANOVA and its graphical results.

## References

- 1 *World Drug Report 2019*, United Nations, 2019, <https://wdr.unodc.org/wdr2019/index.html>.
- 2 *Marijuana Overview*, 2019, <http://www.ncsl.org/research/civil-and-criminal-justice/marijuana-overview.aspx>.
- 3 B. Pearl, *Ending the War on Drugs: By the Numbers*, Center for American Progress, 2018, <https://www.americanprogress.org/issues/criminal-justice/reports/2018/06/27/452819/ending-war-drugs-numbers/>.
- 4 C. Burnier, P. Esseiva and C. Roussel, *Talanta*, 2019, **192**, 135–141.
- 5 S. Zivovinic, R. Alder, M. D. Allenspach and C. Steuer, *J. Anal. Sci. Technol.*, 2018, **9**, 1–10.
- 6 B. Patel, D. Wene and Z. T. Fan, *J. Pharm. Biomed. Anal.*, 2017, **146**, 15–23.



- 7 B. Nie, J. Henion and I. Ryona, *J. Am. Soc. Mass Spectrom.*, 2019, **30**, 719–730.
- 8 K. Leslie, J. Fleischer, S. Jones and J. Piper, *Popular Police Field Tests Can't Tell the Difference Between CBD and Marijuana*, NBC, Washington, 2019, <https://www.nbcwashington.com/news/local/popular-police-field-tests-cant-tell-the-difference-between-cbd-and-marijuana/159828/>.
- 9 D. Kourouski, R. P. Van Duyne and I. K. Lednev, *Analyst*, 2015, **140**, 4967–4980.
- 10 E. Hager, C. Farber and D. Kourouski, *Forensic Chem.*, 2018, **9**, 44–49.
- 11 C. Farber and D. Kourouski, *Anal. Chem.*, 2018, **90**, 3009–3012.
- 12 L. Sanchez, C. Farber, J. Lei, K. Zhu-Salzman and D. Kourouski, *Anal. Chem.*, 2019, **91**, 1733–1737.
- 13 C. Farber, M. Mahnke, L. Sanchez and D. Kourouski, *Trends Anal. Chem.*, 2019, **118**, 43–49.
- 14 H. G. Edwards, D. W. Farwell and D. Webster, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 1997, **53**, 2383–2392.
- 15 N. Tschirner, K. Brose, M. Schenderlein, A. Zouni, E. Schlodder, M. A. Mroginski, P. Hildebrandt and C. Thomsen, *Phys. Status Solidi*, 2009, **246**, 2790–2793.
- 16 K. De Gussem, P. Vandenabeele, A. Verbeken and L. Moens, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 2005, **61**, 2896–2908.
- 17 H. G. M. Edwards, D. W. Farwell and D. Webster, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 1997, **53**, 2383–2392.
- 18 U. P. Agarwal and S. A. Ralph, *Appl. Spectrosc.*, 2016, **51**, 1648–1655.
- 19 U. P. Agarwal, *Front. Plant Sci.*, 2014, **5**, 1–12.
- 20 Y. S. Mary, C. Y. Panicker and H. T. Varghese, *Orient. J. Chem.*, 2012, **28**, 937–941.
- 21 M. M. Yu, H. G. Schulze, R. Jetter, M. W. Blades and R. F. Turner, *Appl. Spectrosc.*, 2007, **61**, 32–37.
- 22 Y. Cao, D. Shen, Y. Lu and J. Huang, *Ann. Bot.*, 2006, **97**, 1091–1094.
- 23 G. Devitt, K. Howard, A. Mudher and S. Mahajan, *ACS Chem. Neurosci.*, 2018, **9**, 404–420.
- 24 F. Adar, *Spectroscopy*, 2017, **32**, 12–20.
- 25 L. Kang, K. Wang, X. Li and B. Zou, *J. Phys. Chem. C*, 2016, **120**, 14758–14766.
- 26 U. P. Agarwal, *Planta*, 2006, **224**, 1141–1153.
- 27 K. Sivashanmugan, K. Squire, A. Tan, Y. Zhao, J. A. Kraai, G. L. Rorrer and A. X. Wang, *ACS Sens.*, 2019, **4**, 1109–1117.
- 28 S. Hoshino, H. Hosoya and S. Nagakura, *Can. J. Chem.*, 1966, **44**, 1961–1965.

